

Signal power amplification of intracellular calcium dynamics with non-Gaussian noises and time delay[☆]



Wei-Long Duan^a, Chunhua Zeng^{b,*}

^a City college, Kunming University of Science and Technology, Kunming 650051, PR China

^b State Key Laboratory of Complex Nonferrous Metal Resources Clean Utilization/Faculty of Science, Kunming University of Science and Technology, Kunming 650093 PR China

ARTICLE INFO

Keywords:

Intracellular calcium oscillation
Reverse resonance
Non-Gaussian noise
Time delay

ABSTRACT

The effect of non-Gaussian noises on stochastic resonance of intracellular Ca^{2+} concentration in intracellular calcium oscillation(ICO) system with time delay is investigated by means of second-order stochastic Runge–Kutta type algorithm. By simulating the signal power amplification(SPA), the results indicate: there are respectively continuous values and a value of the parameter p (which is used to control the degree of the departure from the non-Gaussian noise and Gaussian noise.) to enhance reverse resonance in the behavior of SPA vs. p in cytosol and calcium store, namely continuous reverse resonance occurs in cytosol and reverse resonance occurs in calcium store. Moreover, SPA monotonically decreases as non-Gaussian noises strengthen, and SPA fast decays to constant as correlation time of non-Gaussian noises increases.

© 2016 Published by Elsevier Inc.

1. Introduction

In many studies on ICO, there are a variety of channels showing calcium-induced calcium release and a variety of models to describe ICO [1–4]. Many interesting phenomena have been found such as stochastic resonance [5,6], reverse resonance [6–8], coherence resonance [7], oscillatory coherence [9], resonant activation [10], bistability solutions with hysteresis [11], calcium puffs [12], various spontaneous Ca^{2+} patterns [13], colored noise-optimized calcium wave [14], stochastic backfiring [15], stability transition [16], and dispersion gap and localized spiral waves [17]. More importantly, Matjaž Perc et al. [18–21] has found that noise and other stochastic effects indeed play a central role [18,19] in system. Recently, calcium wave instability [22,23] has also been studied.

Martin Falcke et al. [15,17,24–29] has intensively studied ICO. For instance, a discrete stochastic model for calcium dynamics in living cells [24], spatial and temporal structures in intracellular Ca^{2+} dynamics caused by fluctuations [25], and key characteristics of Ca^{2+} puffs in deterministic and stochastic frameworks [28]. Additionally, they clearly showed that real ICO is non-Gaussian [29]. As stated in above, stochastic resonance and reverse resonance have been discovered in ICO. Thus, in this paper, we study the effect of non-Gaussian noise on stochastic resonance of ICO. For the role of noise on some stochastic systems, there are also some research [30–35].

* This project was supported by the National Natural Science Foundation of China (Grant No. 11305079 and Grant No. 11347014) and Introduction of talent capital group fund project of Kunming University of Science and Technology (Grant No. KKZ3201407030), and the Candidate Talents Training Fund of Yunnan Province (Project No. 2015HB025).

* Corresponding author.

E-mail addresses: yndwl@sina.com.cn (W.-L. Duan), zchh2009@126.com (C. Zeng).

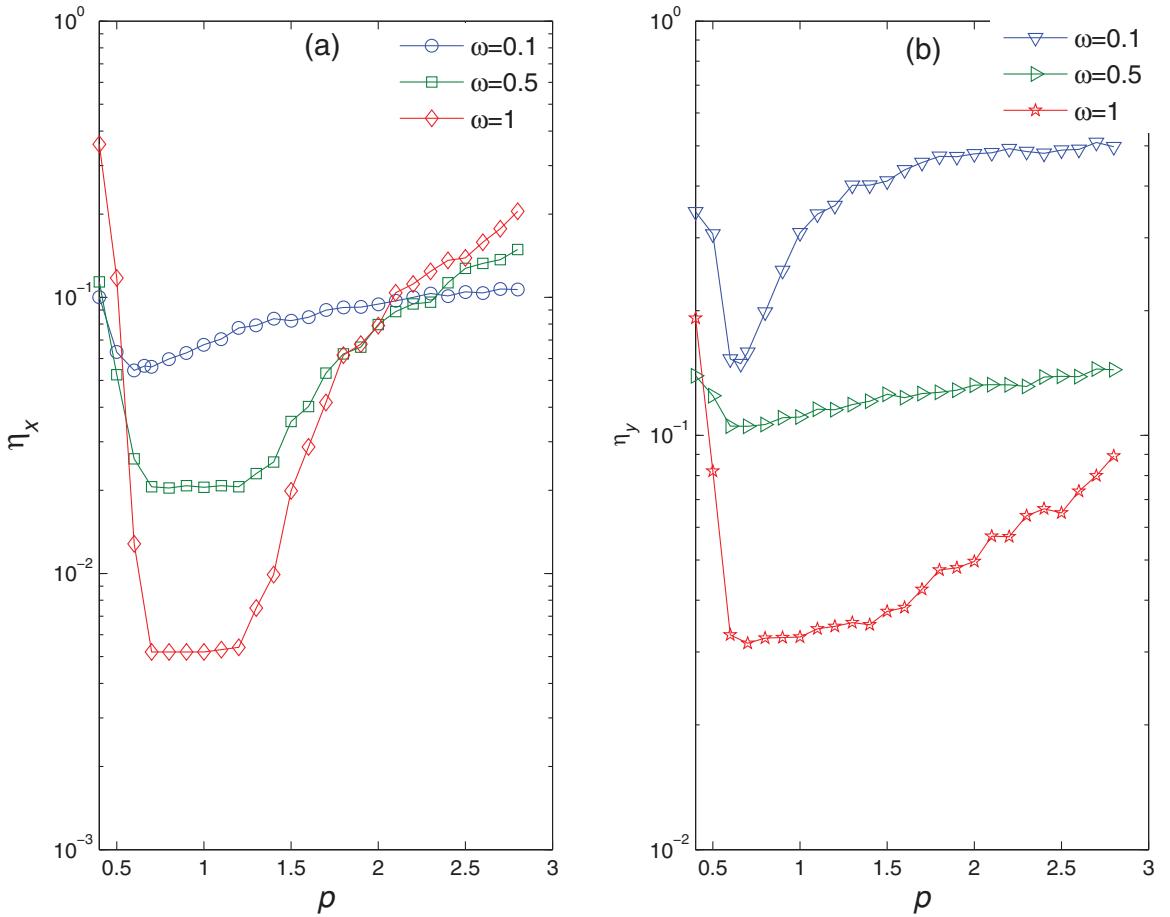


Fig. 1. The SPA η_x (see Fig. 1(a)) and η_y (see Fig. 1(b)) vs. parameter p of non-Gaussian noises.

2. The model for ICO with non-Gaussian noises and time delay

In order to study easily, taking into account same time delay τ in processes of active and passive transport of Ca^{2+} in a real cell. In this paper, x and y denote the concentration of free Ca^{2+} of cytosol and calcium store in a cell, respectively. Based on calcium-induced calcium release, the Langevin equations of ICO system can be read as follows according to our previous results [7,8]:

$$d_t x = A_1(x; x_\tau, y_\tau) + B_1(x; x_\tau, y_\tau) \eta_1(t), \quad (1)$$

$$d_t y = A_2(x, y; x_\tau) + B_2(x, y; x_\tau) \eta_2(t), \quad (2)$$

with

$$A_1(x; x_\tau, y_\tau) = v_0 + v_1 \beta_0 - v_2 + v_{3\tau} + k_f y_\tau - kx, \quad (3)$$

$$A_2(x, y; x_\tau) = v_{2\tau} - v_3 - k_f y, \quad (4)$$

$$B_1(x; x_\tau, y_\tau) = \sqrt{v_1^2 \beta_0^2 + 2v_1 \beta_0 \lambda W + W^2}, \quad (5)$$

$$B_2(x, y; x_\tau) = \sqrt{\frac{v_{2\tau} + v_3 + k_f y}{V}}, \quad (6)$$

$$W(x; x_\tau, y_\tau) = \sqrt{\frac{v_0 + v_1 \beta_0 + v_2 + v_{3\tau} + k_f y_\tau + kx}{V}}, \quad (7)$$

and

$$v_2 = \frac{V_2 x^2}{x^2 + k_1^2}, \quad v_3 = \frac{V_3 x^4 y^2}{(x^4 + k_2^4)(y^2 + k_3^2)}, \quad (8)$$

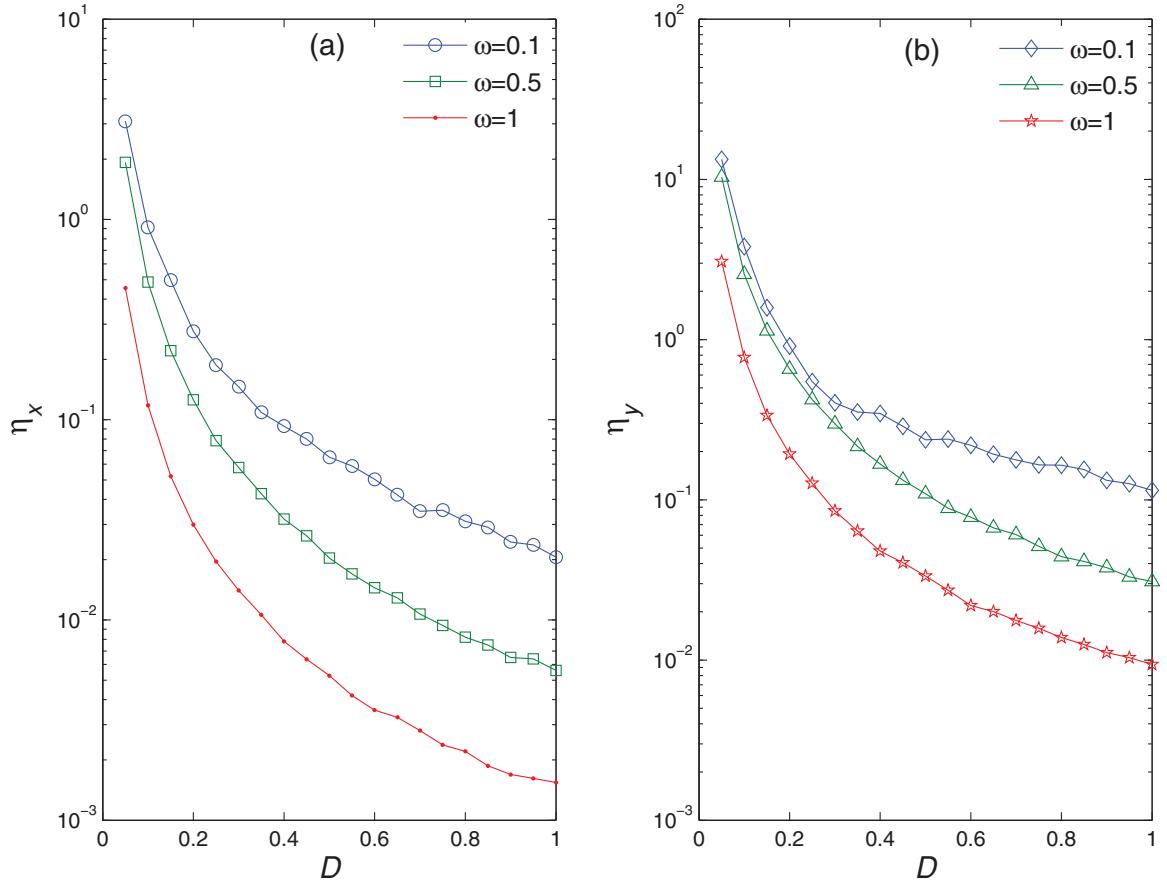


Fig. 2. The SPA η_x (see Fig. 2(a)) and η_y (see Fig. 2(b)) vs. intensity D of non-Gaussian noises.

$$\nu_{2\tau} = \frac{V_2 x_\tau^2}{x_\tau^2 + k_1^2}, \nu_{3\tau} = \frac{V_3 x_\tau^4 y_\tau^2}{(x_\tau^4 + k_2^4)(y_\tau^2 + k_3^2)}. \quad (9)$$

Here ν_0 is the steady flow of Ca^{2+} to the cytosol, ν_1 is the maximum rate of the stimulus induced influx of Ca^{2+} from the extracellular medium, β_0 is the external control parameter that denotes the degree of extracellular simulation. The rates ν_2 and ν_3 refer, respectively, to pumping of Ca^{2+} into calcium store and to release of Ca^{2+} from store into cytosol in a process activated by cytosolic Ca^{2+} . $\nu_{2\tau}$ is ν_2 with time delay, and $\nu_{3\tau}$ is ν_3 with time delay. k_y is a diffusional flow of Ca^{2+} from store to cytosol, k_x denotes the uptake from the cytosol, V is the system size. V_2 and V_3 denote the maximum values of the rates ν_2 and ν_3 , respectively. The parameters k_1 , k_2 , and k_3 are threshold constants for pumping, release, and activation of release by Ca^{2+} and by inositol 1,4,5-trisphosphate. $W = W(x; x_\tau, y_\tau)$, $x_\tau = x(t - \tau)$, $y_\tau = y(t - \tau)$. λ denotes cross-correlation degree of internal and external noise before merger [11].

The noises $\eta_1(t)$ and $\eta_2(t)$ are considered as non-Gaussian noises which are characterized by the following Langevin equation [36]:

$$\frac{d\eta_i(t)}{dt} = -\frac{1}{\tau_1} \frac{d}{d\eta_i} V_{ip}(\eta_i) + \frac{\sqrt{2D}}{\tau_1} \xi_i(t), i = 1, 2. \quad (10)$$

Where $\xi_i(t)$ is a standard Gaussian white noise of zero mean and correlation $\xi_i(t)\xi_i(t') = \delta(t - t').V_{ip}(\eta_i)$ is given by

$$V_{ip}(\eta_i) = \frac{D}{\tau_1(p-1)} \ln \left[1 + \frac{\tau_1}{D}(p-1) \frac{\eta_i^2}{2} \right], \quad (11)$$

and the statistical properties of non-Gaussian noise $\eta_i(t)$ is defined as

$$\langle \eta_i(t) \rangle = 0, \quad (12)$$

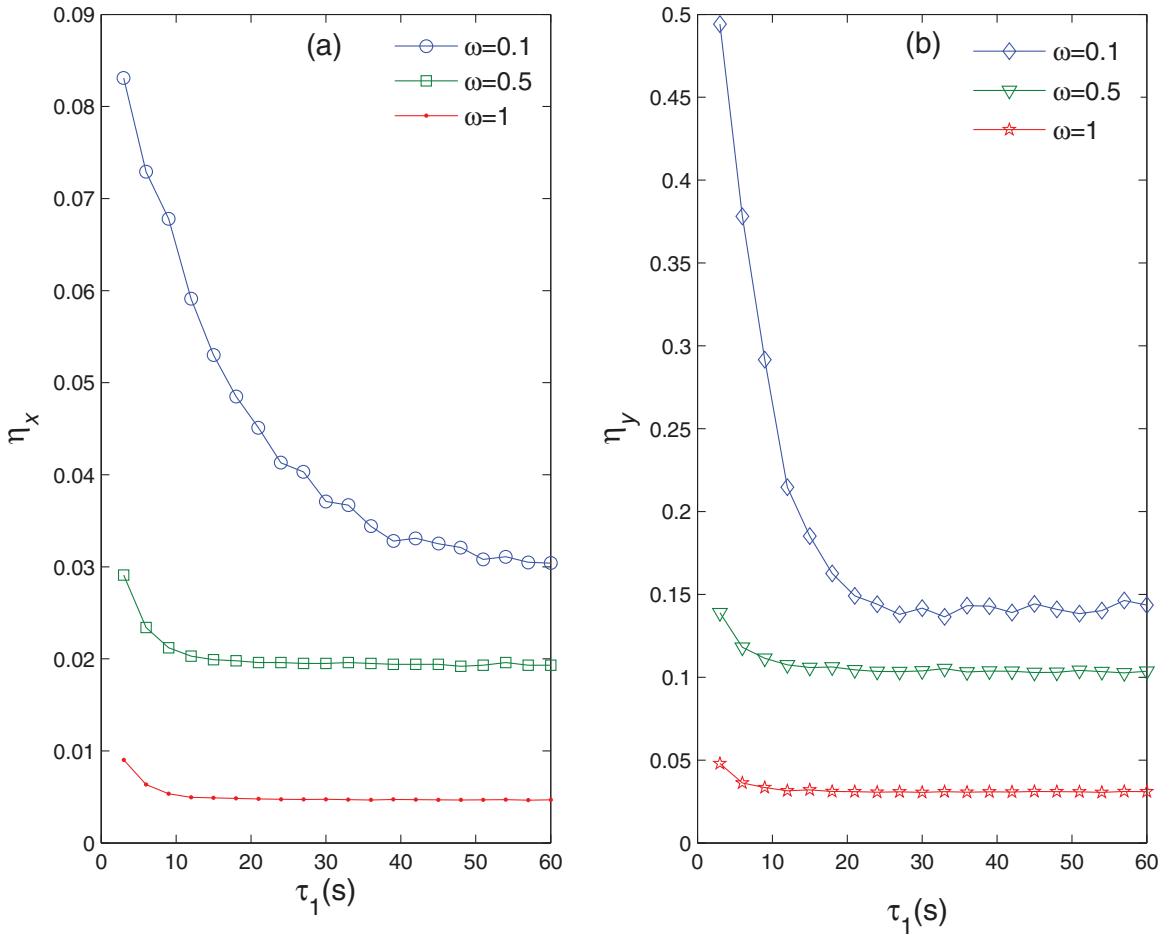


Fig. 3. The SPA η_x (see Fig. 3(a)) and η_y (see Fig. 3(b)) vs. correlation time τ_1 of non-Gaussian noises.

$$\langle \eta_i^2(t) \rangle = \begin{cases} \frac{2D}{\tau_1(5-3p)}, p \in (-\infty, \frac{5}{3}), \\ \infty, p \in [\frac{5}{3}, 3]. \end{cases} \quad (13)$$

Where τ_1 denotes the correlation time of the non-Gaussian noises $\eta_i(t)$, and D denotes the noise intensity of Gaussian white noise $\xi_i(t)$. The parameter p is used to control the degree of the departure from the non-Gaussian noise and Gaussian noise. The distribution of the noise is Gaussian for $p = 1$, non-Gaussian with long tail for $p > 1$, and characterized by a "more than Gaussian" cutoff for $p < 1$. Here, in order to study easily, supposing noises $\xi_1(t)$ and $\xi_2(t)$ have same strength D , and non-Gaussian noises $\eta_1(t)$ and $\eta_2(t)$ have same p and correlation time τ_1 .

3. The signal power amplification

In order to investigate the roles of non-Gaussian noises on stochastic resonance in ICO system, we respectively employ SPA η_x and η_y of cytosolic and calcium store's Ca^{2+} concentration to characterize the stochastic resonance of system [37–39]:

$$\eta_x = 4D^{-2} |\langle e^{i\omega t} X(t) \rangle|, \eta_y = 4D^{-2} |\langle e^{i\omega t} Y(t) \rangle|, \quad (14)$$

where $X(t)$ and $Y(t)$ are obtained respectively from the ensembles average over the stochastic path $x(t)$ and $y(t)$ realizations, and ω is frequency of Fourier transformation. Through integrating Eqs. (1) and (2) with second-order stochastic Runge–Kutta type algorithm [40], the time series $x(t)$ and $y(t)$ can be obtained(for specific simulation algorithm of ICO system, see Ref. [7], discretize time in steps of size $\Delta = 0.001\text{s.}$). After fast Fourier transformation of $X(t)$ and $Y(t)$, one can obtain the amplitude of the first harmonic of the output information η_x and η_y from Eq. (14). By simulating time series, one could obtain many properties of stochastic dynamics system, e.g., in neuronal network system [41–51].

Experimentally, x is in the order of 100–200 nM in basal state [52] and $y = 5 \mu\text{M}$ [53], so that the initial values $x(0)$ and $y(0)$ independently take uniformly random from $0.1 \sim 0.2 \mu\text{M}$ and $4 \sim 5 \mu\text{M}$. In the condition of time delay, it is rational to let $x(t - \tau) = x(0)$ and $y(t - \tau) = y(0)$ as $t < \tau$. In this paper, the value of parameters are set as: $v_0 = 1 \mu\text{M/s}$, $v_1 = 7.3 \mu\text{M/s}$, $\beta_0 = 0.287$, $k_f = 1/\text{s}$, $k = 10/\text{s}$, $V_2 = 65 \mu\text{M/s}$, $V_3 = 500 \mu\text{M/s}$, $k_1 = 1 \mu\text{M}$, $k_2 = 0.9 \mu\text{M}$, $k_3 = 2 \mu\text{M}$, $V = 1000 \mu\text{m}^3$, $\lambda = 0.1$, and $\tau = 0.5 \text{ s}$.

In the following, by simulating SPA η_x and η_y , we respectively discuss the effect of parameter p , intensity D , and correlation time τ_1 of non-Gaussian noises on stochastic resonance in ICO system.

In Fig. 1, the role of parameter p on stochastic resonance is plotted, here $D = 0.5$ and $\tau_1 = 10 \text{ s}$. As shown in figure, the SPA η_x (see Fig. 1(a)) exhibits the deeper valley as ω increases, i.e., the larger the ω , the deeper the valley is. Namely, reverse resonance occurs in cytosol, it enhances as ω increases. It's clear, there is only a value of p to indicate reverse resonance when ω is small (e.g., $\omega = 0.1$). Additionally, as ω increases, the bottom of valley becomes flat (e.g., $\omega = 0.5$ and 1), which demonstrates, the presence of some continuous values of p induce reverse resonance, i.e., reverse resonance always happens as long as p takes the value of in this portion. Briefly, in these continuous values of p , continuous reverse resonance happens in cytosol. However, SPA η_y (see Fig. 1(b)) shows a structure of deep valley with small ω (e.g., $\omega = 0.1$), as ω increases, this structure disappears slowly (e.g. $\omega = 0.5$ and 1), i.e., SPA only presents a minimum with small ω . It shows that reverse resonance only occurs in calcium store when ω is small, as ω increases, reverse resonance decreases.

Then, the role of intensity D of non-Gaussian noises on stochastic resonance is studied in Fig. 2, here $p = 0.9$ and $\tau_1 = 10\text{s}$. It is clearly seen that SPA η_x (see Fig. 2(a)) and η_y (see Fig. 2(b)) linearly decrease as non-Gaussian noises strengthen. Moreover, the higher the ω , the smaller the SPA is.

Finally, the role of correlation time τ_1 of non-Gaussian noises on stochastic resonance is studied in Fig. 3, here $p = 0.9$ and $D = 0.5$. Whether SPA η_x (see Fig. 3(a)) or η_y (see Fig. 3(b)) roughly shows same variation as τ_1 prolongs, i.e., both η_x and η_y fast decay to a constant. Moreover, the larger the ω , the faster the decay is and the smaller the SPA is.

4. Conclusions

In view of non-Gaussian noises in transmission processes of intracellular Ca^{2+} , by means of second-order stochastic Runge–Kutta type algorithm, the effect of non-Gaussian noises on stochastic resonance of intracellular Ca^{2+} concentration in ICO system with time delay is investigated.

By simulating SPA η_x and η_y of cytosolic and calcium store's Ca^{2+} concentration, the results indicate that: (1) in cytosol, there is a value of p to induce reverse resonance with small ω , as ω enlarges continuous reverse resonance occurs at portion continuous values of p , and reverse resonance increases as ω increases; in calcium store, though reverse resonance also occurs, it decreases as ω increases. (2) As non-Gaussian noises strengthen, SPA η_x and η_y show decay all the time, the larger the ω , the smaller the SPA is. (3) As correlation time varies, SPA η_x and η_y fast decay to constant, the larger the ω , the faster the decay is and the smaller the SPA is. Therefore, in ICO system, the value of p is main factor to induce reverse resonance, it may has significance to study ICO and cell biology.

References

- [1] A. Goldbeter, G. Dupont, M.J. Berridge, Minimal model for signal-induced Ca^{2+} oscillations and for their frequency encoding through protein phosphorylation, PNAS 87 (1990) 1461–1465.
- [2] Y. Shiferaw, D. Sato, A. Karma, Coupled dynamics of voltage and calcium in paced cardiac cells, Phys. Rev. E 71 (2005) 021903.
- [3] A.C. Ventura, L. Bruno, S.P. Dawson, Simple data-driven models of intracellular calcium dynamics with predictive power, Phys. Rev. E 74 (2006) 011917.
- [4] R. Thul, M. Falcke, Frequency of elemental events of intracellular Ca^{2+} dynamics, Phys. Rev. E 73 (2006) 061923.
- [5] H. Li, Z. Hou, H. Xin, Internal noise stochastic resonance for intracellular calcium oscillations in a cell system, Phys. Rev. E 71 (2005) 061916.
- [6] W.L. Duan, F. Long, C. Li, Reverse resonance and stochastic resonance in intracellular calcium oscillations, Physica A 401 (2014) 52–57.
- [7] L. Lin, W.L. Duan, The phenomena of an intracellular calcium oscillation system with non-gaussian noises, Chaos, Solitons Fract. 77 (2015) 132–137.
- [8] W.L. Duan, C.H. Zeng, Role of time delay on intracellular calcium dynamics driven by non-gaussian noises, Sci. Rep. 6 (2016) 25067.
- [9] W.L. Duan, Time delay induces oscillatory coherence in intracellular calcium oscillation system, Physica A 405 (2014) 10–16.
- [10] W.L. Duan, P.F. Duan, Time delay induces resonant activation in intracellular calcium oscillations, Chin. J. Phys. 52 (2014) 1059–1068.
- [11] W.L. Duan, L.J. Yang, D.C. Mei, Simulation of time delay effects in the intracellular calcium oscillation of cells, Phys. Scr. 83 (2011) 015004.
- [12] S. Rüdiger, J.W. Shuai, I.M. Sokolov, Law of mass action, detailed balance, and the modeling of calcium puffs, Phys. Rev. Lett. 105 (2010) 048103.
- [13] J.W. Shuai, P. Jung, Selection of intracellular calcium patterns in a model with clustered Ca^{2+} release channels, Phys. Rev. E 67 (2003) 031905.
- [14] W.L. Duan, Z.B. Fan, Colored noises commutes calcium wave in intracellular calcium oscillation, Chin. J. Phys. 52 (2014) 224–232.
- [15] M. Falcke, L. Tsimring, H. Levine, Stochastic spreading of intracellular Ca^{2+} release, Phys. Rev. E 62 (2000) 2636.
- [16] L. Lin, W.L. Duan, Extrinsic periodic information interpolates between monostable and bistable states in intracellular calcium dynamics, Physica A 427 (2015) 155–161.
- [17] M. Falcke, M. Or-Guil, M. Bär, Dispersion gap and localized spiral waves in a model for intracellular Ca^{2+} dynamics, Phys. Rev. Lett. 84 (2000) 4753.
- [18] M. Perc, A.K. Green, C.J. Dixon, M. Marhl, Establishing the stochastic nature of intracellular calcium oscillations from experimental data, Biophys. Chem. 132 (2008) 33–38.
- [19] M. Perc, M. Rupnik, M. Gosak, M. Marhl, Prevalence of stochasticity in experimentally observed responses of pancreatic acinar cells to acetylcholine, Chaos 19 (2009) 037113.
- [20] M. Perc, M. Gosak, M. Marhl, From stochasticity to determinism in the collective dynamics of diffusively coupled cells, Chem. Phys. Lett. 421 (2006) 106–110.
- [21] M. Perc, M. Gosak, M. Marhl, Periodic calcium waves in coupled cells induced by internal noise, Chem. Phys. Lett. 437 (2007) 143–147.
- [22] W.L. Duan, Colored noises decrease correlation in intracellular calcium oscillation, Chin. J. Phys. 52 (2014) 1355–1363.
- [23] C.B. Tabi, I. Maina, A. Mohamadou, H.P.E. Fouda, T.C. Kofané, Wave instability of intercellular Ca^{2+} oscillations, EPL 106 (2014) 18005.
- [24] M. Bär, M. Falcke, H. Levine, L.S. Tsimring, Discrete stochastic modeling of calcium channel dynamics, Phys. Rev. Lett. 84 (2000) 5664.
- [25] R. Thul, M. Falcke, Stability of membrane bound reactions, Phys. Rev. Lett. 93 (2004) 188103.

- [26] K. Thurley, M. Falcke, Derivation of Ca^{2+} signals from puff properties reveals that pathway function is robust against cell variability but sensitive for control, *PNAS* 108 (2011) 427–432.
- [27] M. Falcke, Introduction to focus issue: intracellular Ca^{2+} dynamics a change of modeling paradigm? *Chaos* 19 (2009) 037101.
- [28] R. Thul, K. Thurley, M. Falcke, Toward a predictive model of Ca^{2+} puffs, *Chaos* 19 (2009) 037108.
- [29] K. Thurley, A. Skupin, R. Thul, M. Falcke, Fundamental properties of Ca^{2+} signals, *Biochim. Biophys. Acta* 1820 (2012) 1185–1194.
- [30] X. Wang, Y. Liang, Q. Pan, C. Zhao, F. Yang, Design and implementation of gaussian filter for nonlinear system with randomly delayed measurements and correlated noises, *Appl. Math. Comput.* 232 (2014) 1011–1024.
- [31] X. Wang, J. Duan, X. Li, Y. Luan, Numerical methods for the mean exit time and escape probability of two-dimensional stochastic dynamical systems with non-gaussian noises, *Appl. Math. Comput.* 258 (2015) 282–295.
- [32] C.H. Zeng, C. Zhang, J.K. Zeng, H.C. Luo, D. Tian, H.L. Zhang, F. Long, Y.H. Xu, Noises-induced regime shifts and -enhanced stability under a model of lake approaching eutrophication, *Ecol. Complex.* 22 (2015) 102–108.
- [33] I. Franović, K. Todorović, M. Perc, N. Vasović, N. Burić, Activation process in excitable systems with multiple noise sources: one and two interacting units, *Phys. Rev. E* 92 (2015) 062911.
- [34] C.H. Zeng, C. Zhang, J.K. Zeng, R.F. Liu, H. Wang, Noise-enhanced stability and double stochastic resonance of active brownian motion, *J. Stat. Mech.* (2015) P08027.
- [35] I. Franović, M. Perc, K. Todorović, S. Kostić, N. Burić, Activation process in excitable systems with multiple noise sources: large number of units, *Phys. Rev. E* 92 (2015) 062912.
- [36] L. Borland, Ito-langevin equations within generalized thermostatics, *Phys. Lett. A* 245 (1998) 67–72.
- [37] L. Gammaitoni, P. Hänggi, P. Jung, F. Marchesoni, Stochastic resonance, *Rev. Mod. Phys.* 70 (1998) 223–287.
- [38] P. Jung, P. Hänggi, Stochastic nonlinear dynamics modulated by external periodic forces, *EPL* 8 (1989) 505–510.
- [39] P. Jung, P. Hänggi, Amplification of small signals via stochastic resonance, *Phys. Rev. A* 44 (1991) 8032–8042.
- [40] D. Wu, X.Q. Luo, S.Q. Zhu, Stochastic system with coupling between non-gaussian and gaussian noise terms, *Physica A* 373 (2007) 203–214.
- [41] H.X. Qin, J. Ma, W.Y. Jin, C.N. Wang, Dynamics of electric activities in neuron and neurons of network induced by autapses, *Sci. China Technol. Sci.* 57 (2014) 936–946.
- [42] X.L. Song, C.N. Wang, J. Ma, J. Tang, Transition of electric activity of neurons induced by chemical and electric autapses, *Sci. China Technol. Sci.* 58 (2015) 1007–1014.
- [43] J. Ma, J. Tang, A review for dynamics of collective behaviors of network of neurons, *Sci. China Tech. Sci.* 58 (2015) 2038–2045.
- [44] J. Tang, T.B. Liu, J. Ma, J.M. Luo, X.Q. Yang, Effect of calcium channel noise in astrocytes on neuronal transmission, *Commun. Nonlinear Sci. Numer. Simulat.* 32 (2016) 262–272.
- [45] E. Yilmaz, V. Baysal, M. Perc, M. Ozer, Enhancement of pacemaker induced stochastic resonance by an autapse in a scale-free neuronal network, *Sci. China Tech. Sci.* 59 (2016) 364–370.
- [46] J. Ma, X. Song, W. Jin, C. Wang, Autapse-induced synchronization in a coupled neuronal network, *Chaos, Solitons Fract.* 80 (2015) 31–38.
- [47] Y. Wang, Z. Wang, J. Wang, Y. Wang, Z. Henderson, X. Wang, X. Zhang, J. Song, C. Lu, The modulation of nicotinic acetylcholine receptors on the neuronal network oscillations in rat hippocampal CA3 area, *Sci. Rep.* 5 (2015) 09493.
- [48] L. Ford, M. Crossley, T. Williams, J.R. Thorpe, L.C. Serpell, G. Kemenes, Effects of $\alpha\beta$ exposure on longterm associative memory and its neuronal mechanisms in a defined neuronal network, *Sci. Rep.* 5 (2015) 10614.
- [49] J. Ma, X. Song, J. Tang, C. Wang, Wave emitting and propagation induced by autapse in a forward feedback neuronal network, *Neurocomputing* 167 (2015) 378–389.
- [50] A. Bikbaev, R. Frischknecht, M. Heine, Brain extracellular matrix retains connectivity in neuronal networks, *Sci. Rep.* 5 (2015) 14527.
- [51] X. Song, C. Wang, J. Ma, J. Tang, Transition of electric activity of neurons induced by chemical and electric autapses, *Sci. China Tech. Sci.* 58 (2015) 1007–1014.
- [52] J.B. Hoek, J.L. Farber, A.P. Thomas, X. Wang, Calcium ion-dependent signalling and mitochondrial dysfunction: mitochondrial calcium uptake during hormonal stimulation in intact liver cells and its implication for the mitochondrial permeability transition, *Biochim. Biophys. Acta* 1271 (1995) 93–102.
- [53] A.D. Short, M.G. Klein, M.F. Schneider, D.L. Gill, Inositol 1,4,5-trisphosphate-mediated quanta Ca^{2+} release measured by high resolution imaging of Ca^{2+} within organelles, *J. Biol. Chem.* 268 (1993) 25887–25893.